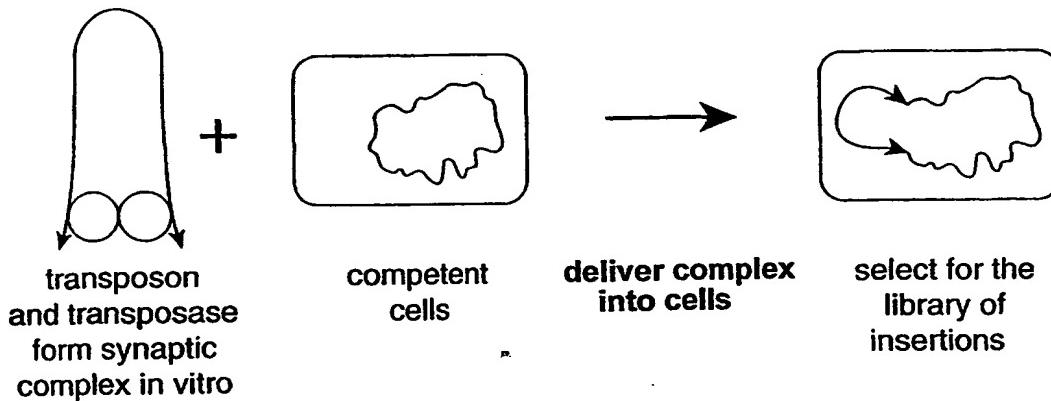




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(54) Title: METHOD FOR MAKING INSERTIONAL MUTATIONS

Synaptic complex transformation

(57) Abstract

A method for making insertional mutations at random or quasi-random locations in the chromosomal or extra-chromosomal nucleic acid of a target cell includes the step of combining, in the target cell, cellular nucleic acid with a synaptic complex that comprises: (a) a Tn5 transposase protein and (b) a polynucleotide that comprises a pair of nucleotide sequences adapted for operably interacting with Tn5 transposase and a transposable nucleotide sequence therebetween, under conditions that mediate transpositions into the cellular DNA. In the method, the synaptic complex is formed *in vitro* under conditions that disfavor or prevent the synaptic complexes from undergoing productive transposition.

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METHOD FOR MAKING INSERTIONAL MUTATIONS

CROSS-REFERENCE TO RELATED APPLICATIONS

Not applicable.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH
5 OR DEVELOPMENT

Not applicable.

BACKGROUND OF THE INVENTION

Efficient insertion of exogenous nucleic acid into the chromosomal and extra-chromosomal nucleic acid of cells is 10 desired in the art of molecular biology to identify chromosomal regions involved in expressing or regulating expression of peptides and proteins. This same technology is also advantageously used in developing new therapeutic and pharmacologic agents.

15 One common method relies upon *in vivo* Tn5 mutagenesis to insert polynucleotides of interest into cellular DNA and to construct libraries of cells that contain inserted polynucleotides at random or quasi-random locations. Existing *in vivo* Tn5 mutagenesis methods require target 20 cells to encode transposase, either natively or from an introduced expression construct. Accordingly, it can be necessary to construct a suitable expression system appropriate to each target cell type. This can be time consuming, and requires extensive knowledge of the 25 requirements of each target cell type.

In many cases, the gene that encodes transposase is encoded by an active transposon, which can continue to transpose in a target cell after the initial desired mutagenesis step. Such undesired residual transposition is 30 undesired in that it complicates the analysis of insertional mutant libraries.

Furthermore, many techniques for *in vivo* Tn5 mutagenesis rely upon a complex biological mechanism for introducing exogenous DNA into the target cells, such as bacteriophage lambda transducing phage or a conjugating 5 plasmid. It would be desirable to avoid requiring such complex biological systems.

Shoji-Tanaka, A., et al., B.B.R.C. 203:1756-1764 (1994) describe using purified retroviral integrase to mediate gene transfer into murine cells.

10 Kuspa, A. and W.F. Loomis, P.N.A.S. U.S.A. 89:8803-8807 (1992) and others have described specifically integrating a plasmid linearized with a restriction enzyme into a genomic restriction site by electroporating enzyme-cut nucleic acid along with the cleaving enzyme into target 15 cells.

BRIEF SUMMARY OF THE INVENTION

The present invention is summarized in that a method for efficiently inserting a transposable polynucleotide at random or quasi-random locations in the chromosomal or 20 extra-chromosomal nucleic acid of a target cell includes the step of combining, in the target cell, cellular nucleic acid with a synaptic complex that comprises (a) a Tn5 transposase protein complexed with (b) a polynucleotide that comprises a pair of nucleotide sequences adapted for 25 operably interacting with Tn5 transposase and a transposable nucleotide sequence therebetween, under conditions that mediate transpositions into the cellular DNA. In the method, the synaptic complex is formed *in vitro* under conditions that disfavor or prevent the 30 synaptic complexes, which are poised for transposition, from actually undergoing productive transposition. The frequency of productive transposition of the transposable nucleotide sequence into the target nucleic acid can be enhanced by using in the method either a hyperactive 35 transposase or a transposable polynucleotide that comprises sequences particularly well adapted for efficient

transposition in the presence of Tn5 transposase, or both.

The present invention is further summarized in that a method for forming library of cells that comprise insertional mutations includes the steps of combining in a 5 plurality of target cells the cellular nucleic acid with the synaptic complex as described, and screening for cells that comprise insertional mutations.

In another aspect, the invention is further summarized as a library of cells that comprise insertional mutations 10 formed according to the above-mentioned method. Such populations of cells that comprise random and independent mutational insertions in their genomes can be screened to select those cells that comprise an insertional mutation that induces a phenotypic or genotypic change relative to 15 cells that were not subject to insertional mutagenesis.

It is an advantage of the present invention that the transposable polynucleotides used to form synaptic complexes can consist of transposon DNA apart from any flanking sequences. This is advantageous in that it 20 reduces the likelihood of intramolecular transposition and increases the likelihood of transposition into a target genome. Moreover, eliminating donor backbone (DBB) sequences from the polynucleotide simplifies preparation of the transposon sequences that can be used in the method.

25 It is another advantage of the present invention that the synaptic complex can form under conditions that disfavor non-productive intramolecular transposition events. This is advantageous in that substantially all of the synaptic complexes can undergo transposition when 30 combined with the cellular DNA. Little, if any, of the nucleic acid in the synaptic complexes is inactive.

It is a feature of the present invention that transposition-promoting conditions are encountered only after the synaptic complex is in the presence of the target 35 nucleic acid in the target cell.

Other objects, advantages, and features of the present invention will become apparent upon consideration of the

following detailed description.

BRIEF DESCRIPTION OF THE SEVERAL VIEWS OF THE DRAWINGS

Fig. 1 depicts the transfer of a synaptic complex into a target cell followed by selection of insertion mutants.

5 Fig. 2 a preferred transposase enzyme for use in the method.

Fig. 3 depicts transposon terminal sequences that can be used in the transposable polynucleotide sequence in the present invention.

10 Fig. 4 depicts a transposon or transposable polynucleotide sequence having mosaic ends and a transposable nucleic acid sequence therebetween.

DETAILED DESCRIPTION OF THE INVENTION

In a system for *in vitro* transposition using modified 15 Tn5 transposase described in International Application No. PCT/US97/15941 (International Publication No. WO 98/10077), incorporated herein by reference in its entirety, the applicants demonstrated productive transposition of a transposable polynucleotide *in vitro* using a modified 20 mutant Tn5 transposase and modified Tn5 polynucleotide sequences flanking a transposable nucleotide sequence.

The present invention differs from the prior system in that in the present invention synaptic complexes are formed *in vitro*, although *in vitro* transposition is substantially 25 absent. Rather, the synaptic complexes are introduced into target cells whereupon transposition *in vivo* readily occurs. Fig. 1 presents a schematic view of one aspect of the present Tn5-based method for efficiently introducing 30 insertional mutations at random or quasi-random locations in the chromosomal or extra-chromosomal nucleic acid of a target cell. In the method, cellular nucleic acid in a target cell is combined with a synaptic complex that comprises (a) a Tn5 transposase protein (shown as a pair of adjacent circles in Fig. 1) and (b) a polynucleotide that 35 comprises a pair of inverted nucleotide sequences (shown as

arrows in Fig. 1) adapted for operably interacting with Tn5 transposase and a transposable nucleotide sequence therebetween, under conditions that mediate transpositions into the cellular DNA. The synaptic complex structure is 5 an intermediate in the transposition of the transposable polynucleotide into the cellular nucleic acid. It is herein disclosed that one can separate synaptic complex formation from transposition by interfering with the ability of the synaptic complex to participate in a polynucleotide strand 10 transfer reaction. In the method, the synaptic complex is formed *in vitro* under conditions that disfavor or prevent the synaptic complexes from undergoing productive transposition. The invention can expand the use of Tn5 mutagenesis methods to, for example, bacteria that are not 15 close relatives of *E. coli* and for which little is known about the structures of, and other requirements for, promoters and other regulatory sequences. In such cells, transposase may not be expressed in adequate amount, or may be unstable. The present method avoids the need to produce 20 an endogenous transposase molecule. Moreover, since the introduced transposase is integrally complexed with the transposon polynucleotide, it is poised to act in the presence of any target DNA and, as such is sufficiently stable to facilitate transposition of the polynucleotide 25 with which it is complexed. One method discovered by the inventors to prevent the synaptic complexes from undergoing polynucleotide strand transfer is achieved by reducing or eliminating magnesium ions (Mg^{++}) from the reaction mixture. Thus, in a preferred embodiment of the method, a suitable 30 transposase and a suitable transposable DNA element are combined in an *in vitro* reaction where the reaction mixture contains magnesium at a level insufficient to support polynucleotide strand transfer, or more preferably, a mixture free of magnesium ions. A suitable reaction buffer 35 for forming the synaptic complexes can be the reaction buffer described on page 19 of the incorporated international application, modified to remove the magnesium

acetate. One can also remove the BSA and spermidine from the reaction buffer. The tRNA can be eliminated from the buffer, unless nucleases are present in the reaction, in which case the tRNA can be added. It may be possible to 5 further simplify the reaction buffer. A typical reaction is described below in the example.

The synaptic complexes are poised to mediate efficient transposition and can be advantageously placed in cold storage until needed. The synaptic complexes can be 10 introduced into suitable target cells capable of providing conditions favorable for transposition, whereupon efficient transposition occurs. By introducing the synaptic complexes into a plurality of suitable target cells and selecting those cells that comprise insertional mutations, 15 a library of cells that comprise random or quasi-random insertional mutations can be produced. To prepare a library of insertional mutants, the transposable nucleic sequence preferably contains a convenient selectable marker such as a gene that confers antibiotic resistance, so that cells 20 lacking an insertional mutation in their cellular DNA can be readily distinguished from those in which productive transposition has occurred. Libraries formed according to the method can be screened for genotypic or phenotypic changes after transposition. At the molecular genetic 25 level, one can employ standard analytical methods including hybridization, restriction fragment mapping, nucleotide sequencing, and combinations thereof, or other methods to identify genetic changes. At the phenotypic level, one can evaluate members of a library of transposition mutants for 30 individual mutants having an altered growth property or other phenotype.

A kit of synaptic complexes comprising transposable polynucleotide sequences having desired characteristics, such as those described below, can be commercialized to 35 facilitate rapid preparation of libraries comprising insertional mutations, wherein the inserted sequences have been designed to accomplish a particular goal, as described

below. The synaptic complexes of the present invention are substantially free of polynucleotide molecules that have undergone productive transposition. Furthermore, because the synaptic complexes are prepared in an *in vitro* reaction, the complexes can be provided as a substantially pure preparation apart from other proteins, genetic material, and the like.

The Tn5 transposase in the synaptic complexes can be a Tn5 transposase that forms synaptic complexes efficiently ¹⁰ *in vitro* (e.g., about 25% or more of the transposon DNA is converted into synaptic complexes with the transposase). The transposase can be a hyperactive Tn5 transposase such as that disclosed in International Application No.

PCT/US97/15941. A preferred mutant Tn5 transposase is a ¹⁵ mutant Tn5 transposase modified relative to a wild-type Tn5 transposase, the mutant transposase comprising a mutation at position 54, and a mutation at position 372, the mutant transposase having greater avidity for Tn5 outside end repeat sequences of a donor DNA and a lesser ability to ²⁰ form multimers than wild-type Tn5 transposase. A mutation at position 54 that confers a greater avidity for Tn5 outside end repeat sequences is a mutation from wild-type glutamic acid to lysine. A mutation at position 372 that causes a reduced ability to form non-productive multimers ²⁵ is a mutation from wild-type lysine to proline.

It is also preferred that the transposase be free of the so-called inhibitor protein, a protein encoded in partially overlapping sequence with the transposase which can interfere with transposase activity. In the method, ³⁰ the transposase is used in purified, or partially purified, form, and if the transposase enzyme is obtained from cells (using conventional methods) it may be possible to separate the transposase from the inhibitor protein before use in the method. However, it is also possible to genetically ³⁵ eliminate the possibility of having any contaminating inhibitor protein present by simply removing its start codon from the gene that encodes the transposase.

An AUG in the wild-type Tn5 transposase gene that encodes methionine at transposase amino acid 56 is the first codon of the inhibitor protein. However, it has already been shown that replacement of the methionine at 5 position 56 has no apparent effect upon the transposase activity, but at the same time prevents translation of the inhibitor protein, thus resulting in a somewhat higher transposition rate. Wygand, T. W. and W. S. Reznikoff, "Characterization of Two Hypertransposing Tn5 Mutants," J. Bact. 174:1229-1239 (1992), incorporated herein by reference. In particular, the present inventors have replaced the methionine with an alanine in the preferred embodiment (and have replaced the methionine-encoding AUG codon with an alanine-encoding GCC). A preferred 10 transposase of the present invention therefore includes an amino acid other than methionine at amino acid position 56, although this change can be considered merely technically advantageous (since it ensures the absence of the inhibitor protein from the *in vitro* system) and not essential to the 15 invention (since other means can be used to eliminate the inhibitor protein from the *in vitro* system). Shown schematically in Fig. 2 is a preferred transposase enzyme having mutations at positions 54, 56 and 372 relative to wild type Tn5 transposase.

The transposable polynucleotide in the synaptic complexes is characterized as a nucleotide sequence flanked by a pair of inverted polynucleotides that comprise an 18 or 19 base long sequence that can function in an *in vitro* Tn5 transposition system. The polynucleotide (or any 20 portion thereof) can be synthesized using methods well known to those skilled in the art, or can be prepared using methods for genetically engineering nucleic acid fragments. Referring now to Fig. 3, the known nucleic acid sequence of 25 an outside end (OE) terminal sequence of a wild-type Tn5 transposon sequence (5'-CTGACTCTTATACACAAGT-3') can be used as the flanking polynucleotides, as can a terminus of an 30 inside end (IE) terminal sequence of a wild-type Tn5

transposon sequence (5'-CTGTCTCTTGATCAGATCT-3'). Although the wild-type OE terminus sequence can be used, the inventors have demonstrated that transposition frequencies at least as high as, and typically significantly higher than, that of wild-type OE can be achieved if the termini in a construct are mosaics intermediate between OE and IE sequences. Preferred mosaic terminus sequences include - bases ATA at positions 10, 11, and 12, respectively, as well as the nucleotides in common between wild-type OE and IE (e.g., at positions 1-3, 5-9, 13, 14, 16, and optionally 19). The nucleotides at position 4, 15, 17, and 18 can correspond to the nucleotides found in those positions in either wild-type OE or wild-type IE. It is noted that the transposition frequency can be enhanced over that of wild-type OE if the nucleotide at position 4 is a T, when in combination with the above-noted nucleotides. Combinations of non-identical ends can also be employed.

The transposable polynucleotide is preferably a linear polynucleotide comprising at its termini the inverted polynucleotide sequences required for Tn5 transposition. It is also possible, but less preferred, to employ a transposable polynucleotide that is linear but which comprises sequences other than the transposable nucleic acid and the inverted polynucleotide sequences. Such additional sequences can be located to either side of the transposable polynucleotide, although this arrangement is less preferred because it complicates preparation of transposon DNA. Some transposition can occur when the transposable polynucleotide is provided on a circular, supercoiled DNA molecule, although the transposition frequency is not as high as when the transposable sequence is a linear molecule.

The transposable nucleic acid sequence between the required end sequences can include any sequence that is desirably inserted into a target genome. Fig. 4 shows a schematic transposon having so-called mosaic ends and a transposable polynucleotide sequence therebetween. One

skilled in the art can readily construct a desired transposable nucleic acid sequence to accomplish a particular goal. The invention is not limited to any particular nucleic acids between the inverted 5 polynucleotide sequences. Rather, the sequence can be any detectable sequence, or a sequence that encodes a product that can be detected using methods known to those skilled in the art. By way of non-limiting example, the transposable nucleic acid sequence can provide the target 10 cell with a selectable marker, which can be a peptide or protein encoded by the transposable nucleic acid sequence. The sequence can encode a protein that confers antibiotic resistance to a cell. Alternatively, the transposable polynucleotide can comprise a sequence whose presence can 15 be detected in target cells. Such a sequence can include a cleavage sequence for a rare restriction enzyme, or any sequence for which a probe exists.

The transposable polynucleotide can also include sequences that regulate expression (transcription or 20 translation) of nearby sequence. The regulatory sequences can facilitate expression of a protein or peptide encoded by a coding sequence in the transposable polynucleotide. Alternatively, the transposable polynucleotide can include regulatory sequences apart from any coding sequence or 25 coding sequences apart from any regulatory sequence. In the former case, transposition events upstream from genomic coding sequences can provide the regulatory elements necessary to modulate transcription and/or translation of an endogenous coding sequence. In the later case, the 30 transposed sequence can reveal previously unknown regulatory sequences in the genome by introducing a marker gene whose product is produced only when the gene is transposed to a position adjacent to a regulatory sequence in the cellular nucleic acid.

35 Once formed from the above-identified components *in vitro*, the synaptic complex can be introduced into target cells using methods known to those of ordinary skill in the

art. The synaptic complexes can alternatively be stored in advance of use, for example, at -40°C. A preferred method for introducing the synaptic complex into the cell is electroporation, for example, using the example of Dauer,
5 W.J. et al, Nucleic Acids Research, 16:6127 (1988), incorporated herein by reference in its entirety. Transposition follows delivery into the cellular nucleic acid without additional intervention. If the synaptic complexes are present in a volume greater than a few
10 microliters, it is also preferred to dialyze the synaptic complexes against a low-salt buffer, e.g., without limitation, 6 mM Tris, pH 7.5 with 7-10% glycerol, before use in an electroporation reaction to reduce salts in the reaction mixture to a level sufficiently low that they do
15 not cause arcing between the electrodes. Other suitable methods for introducing the synaptic complexes into target cells are known and can include transformation, transfection, and liposome-mediated methods.

The system provides a method for introducing mutations into cells quickly and efficiently. Because the transposable polynucleotide sequence is provided, the system is not host-specific, and should work in any target cell. The invention finds particular utility in cells, particularly bacterial cells, that do not encode
25 transposase, since the required transposase molecule is provided directly as part of the synaptic complex. The method is demonstrated to work in non-nucleated target systems, such as bacterial cells. In particular, the synaptic complexes have been introduced into *E. coli* cells
30 (strain MG1655) and productive transposition has been observed. No scientific impediment is known to exist that would prevent use of the method in nucleated cells, such as archaebacteria, plant and animal cells, especially cells of nematodes, amphibians, and mammals, including, but not
35 limited to, rodents and humans. In methods for introducing synaptic complexes into nucleated cells, it may be preferable to provide a nuclear localization signal on the

synaptic complex, preferably as part of a genetically modified transposase protein.

The present invention will be more readily understood upon consideration of the following example which is 5 exemplary and is not intended to limit the scope of the invention.

EXAMPLE

In a reaction of 1 μ l, 0.05 μ g of a purified hyperactive transposase (EK45/MA56/LP372), the amino acid 10 sequence of which is reported in incorporated International Application No. PCT/US97/15941, was combined with 0.1 μ g of a transposable polynucleotide comprising an expression cassette that encodes a protein that confers kanamycin resistance upon a target cell. The expression cassette was 15 flanked by the mosaic ends described in the same incorporated International Application and shown in Fig. 3. The polynucleotide was provided, in separate reactions, as a supercoiled plasmid, a linearized plasmid, or as a polynucleotide fragment comprising at its termini inverted 20 sequences required for transposition mediated by Tn5 transposase.

The mixture was incubated for one hour in the presence or absence of magnesium ions (Mg^{++}) in the reaction buffer noted above. In the absence of magnesium ions, synaptic 25 complexes form, but no transposition occurs *in vitro*.

After incubation, the reaction mixture was mixed with 40 μ l of *E. coli* strain MG1655 and subjected to electroporation according to the incorporated method of Dauer. The cells (1.4×10^9) were plated on LB-Kan plates 30 and kanamycin resistant colonies were counted. When 10 μ l of reaction mix was used, the mix was dialyzed into a suitable buffer to prevent arcing between the electrodes.

Table 1. Tn5 Synaptic Complex
electroporation/transposition

DNA	Tnp	Mg ⁺⁺	volume	Kan ⁴ CFU
supercoiled*	-	+	1 μl	None
5 supercoiled*	+	+	1 μl	1.6x10 ³
linearized*	+	+	1 μl	6.9x10 ³
released element (PvuII cut)	+	+	1 μl	3.9x10 ³
released element	+	-	1 μl	5.3x10 ⁴
10 released element	+	-	1 μl (dialyzed)	4.0x10 ⁴
released element	+	-	10 μl (dialyzed)	5.5x10 ⁵

*contained donor backbone sequences

It is apparent from the table that transposition of the transposable polynucleotide that confers kanamycin resistance upon a cell is high when the reaction mixture is incubated in the absence of magnesium ions. Superior transposition was observed when the transposable polynucleotide contains no sequences flanking the transposable portion ("released element"). A greater than 10 fold reduction in kanamycin resistant CFUs was observed when magnesium was added to the reaction mixture, presumably as a result of non-productive intramolecular transposition that can occur in the presence of magnesium ions.

25 In constructing a library, to achieve a 99% probability of complete coverage (i.e., an insert into each ORF) approximately 20,000 transposition events are required. The assumptions in this calculation are that transposition is truly random with respect to ORF target choice and that each ORF is the same size. Although Tn5 inserts have been found in all genes for which they have been sought, Tn5 does have target sequence biases that can

skew the randomness in terms of gene distribution. There may also be unknown biases imposed on transposition by the condensation/organization of the nucleoid body. In addition, all ORFs are not the same size, but range in size 5 in *E. coli*, for example, from under 100 codons to 2,383 codons. On the other hand, the results of synaptic complex electroporation/transposition described above represent - inserts into dispensable functions only, since only viable colony forming units are formed. This clearly under 10 represents the entire transposition insertion pool. Accordingly, to form a fully representative library, pools of between 50,000 and 500,000 viable transposition events should be sought. The invention described herein is sufficiently efficient to produce a library of this size. 15 The present invention is not intended to be limited to the foregoing, but to encompass all such variations and modifications as come within the scope of the appended claims.

CLAIMS

WE CLAIM:

1. A method for making an insertional mutation at a random or quasi-random position in cellular nucleic acid in 5 a target cell, the method comprising the step of:

introducing into the target cell a synaptic complex - that comprises (a) a Tn5 transposase protein and (b) a polynucleotide that comprises a pair of nucleotide sequences adapted for operably interacting with Tn5 10 transposase to form a synaptic complex and a transposable nucleotide sequence therebetween, under conditions that mediate transpositions into the cellular nucleic acid.

2. A method as claimed in Claim 1 wherein the method further comprises the step of:

15 combining the Tn5 transposase protein and the polynucleotide *in vitro* under conditions that disfavor polynucleotide strand transfer to form the synaptic complex.

3. A method as claimed in Claim 2 wherein the Tn5 20 transposase protein and the polynucleotide are combined *in vitro* in a reaction that comprises magnesium ions at a level insufficient to support polynucleotide strand transfer.

4. A method as claimed in Claim 3 wherein the 25 reaction lacks magnesium ions.

5. A method as claimed in Claim 1 wherein the Tn5 transposase is a mutant Tn5 transposase modified relative to a wild type Tn5 transposase, the mutant transposase comprising:

5 a mutation at position 54; and
a mutation at position 372,
the mutant transposase having greater avidity for Tn5 outside end repeat sequences of a donor DNA and a lesser ability to form multimers than wild type Tn5 transposase.

10 6. A method as claimed in Claim 5 wherein the mutation at position 54 is a substitution mutation.

7. A method as claimed in Claim 6, wherein position 54 is a lysine.

8. A method as claimed in Claim 5 wherein the 15 mutation at position 372 is a substitution mutation.

9. A method as claimed in Claim 8 wherein position 372 is a proline.

10. A method as claimed in Claim 5 wherein the Tn5 transposase further comprises a substitution mutation at 20 position 56, wherein the mutant transposase lacks an inhibitor activity.

11. A method as claimed in Claim 10 wherein position 56 is alanine.

12. A method as claimed in Claim 1 wherein the nucleotide sequence adapted for operably interacting with Tn5 transposase is an 18 or 19 base pair polynucleotide sequence that comprises nucleotide A at position 10, 5 nucleotide T at position 11, and nucleotide A at position 12.

13. A method as claimed in Claim 12 wherein the nucleotide sequence has the sequence 5'-
CTGTCTCTTATACACATCT-3'.

10 14. A method as claimed in Claim 12 wherein the nucleotide sequence has the sequence 5'-
CTGTCTCTTATACAGATCT-3'.

15. A method for forming a synaptic complex between
15 (a) a Tn5 transposase protein and (b) a polynucleotide that
comprises a pair of nucleotide sequences adapted for
operably interacting with Tn5 transposase to form a
synaptic complex and a transposable nucleotide sequence
therebetween, the method comprising the step of combining
10 (a) and (b) *in vitro* under conditions that disfavor
polynucleotide strand transfer to form the synaptic
complex.

16. A method as claimed in Claim 15 wherein the Tn5
transposase protein and the polynucleotide are combined *in*
vitro in a reaction that comprises magnesium ions at a
25 level insufficient to support polynucleotide strand
transfer.

17. A method as claimed in Claim 16 wherein the reaction lacks magnesium ions.

18. A method for forming a library of insertional mutations at random or quasi-random positions in cellular nucleic acid in a plurality of target cells, the method comprising the steps of:

introducing into the target cells a synaptic complex that comprises (a) a Tn5 transposase protein and (b) a polynucleotide that comprises a pair of nucleotide sequences adapted for operably interacting with Tn5 transposase to form a synaptic complex and a transposable nucleotide sequence therebetween, the transposable nucleotide sequence comprising a selectable marker, under conditions that mediate transpositions into the cellular nucleic acid; and

screening for the cells that comprise the selectable marker.

1/2

Synaptic complex transformation

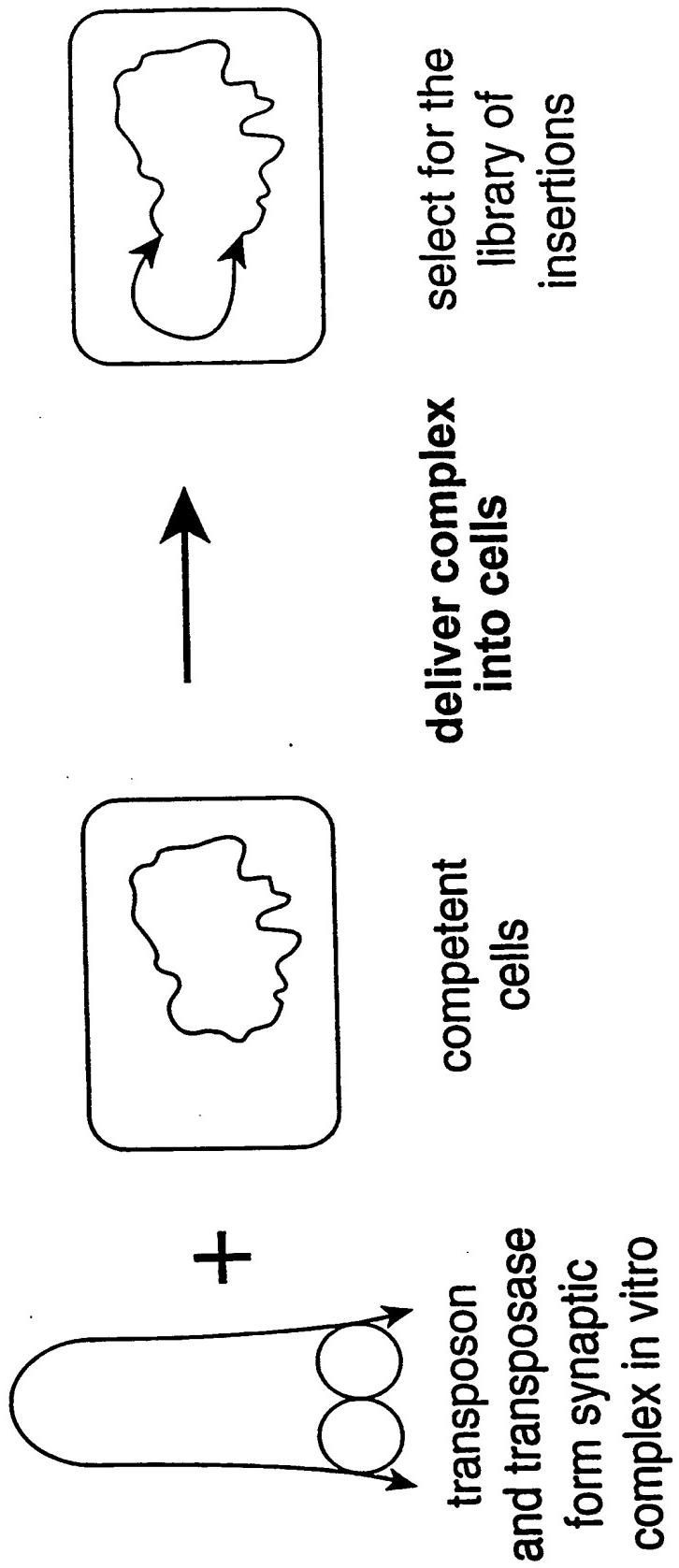
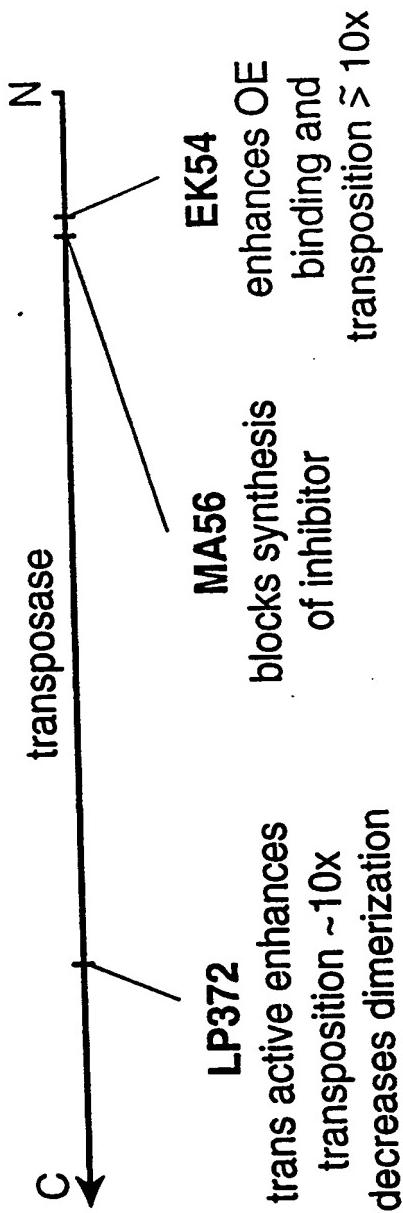


FIG 1

2/2

FIG 2 Tmp (EK54, MA56, LP372)



3
FIG

19
 OE MOSAIC IE
 1 C T G A C T C T T A T A C A C A A G T
 C T G T C T C T T A T A C A C A T C T
 C T G T C T C T T G A T C A G A T C T

FIG 4

ME transposable nucleic acid

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 99/21960

A. CLASSIFICATION OF SUBJECT MATTER
 IPC 7 C12N15/10 C12N15/55 C12N15/90 C12N9/22

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
 IPC 7 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	GORYSHIN I Y ET AL.: "Tn5 in vitro transposition" JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 273, no. 13, 27 March 1998 (1998-03-27), pages 7367-7374, XP002129603 page 7371, left-hand column, line 10-14	15-17
Y	SAKAI J ET AL.: "Identification and characterization of a pre-cleavage synaptic complex that is an early intermediate in Tn10 transposition" EMBO JOURNAL, vol. 14, no. 17, 1 September 1995 (1995-09-01), pages 4374-4384, XP002129605 page 4377, left-hand column, line 16-25; figure 5	15-17

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
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- "O" document referring to an oral disclosure, use, exhibition or other means
- "T" document published prior to the International filing date but later than the priority date claimed

"T" later document published after the International filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

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Date of the actual completion of the International search

10 February 2000

Date of mailing of the International search report

28/02/2000

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 98 10077 A (WISCONSIN ALUMNI RES FOUND) 12 March 1998 (1998-03-12) cited in the application claims 1-14 page 7, line 5-30 page 19, line 5-19	1,2, 5-15,18
A	PARK, BYUNG TAE ET AL: "In vitro transposition of Tn5" TAEHAN MISAENGMUL HAKHOECHI (JOURNAL OF THE KOREAN SOCIETY FOR MICROBIOLOGY), vol. 27, no. 4, 1992, pages 381-389, XP000874603 page 386, paragraph 6; table 3	1-4, 15-17
A	JUNOP M S ET AL.: "Multiple roles for divalent metal ions in DNA transposition: distinct stages of Tn10 transposition have different Mg ²⁺ requirements." EMBO JOURNAL, vol. 15, no. 10, 15 May 1996 (1996-05-15), pages 2547-2555, XP002129619 abstract	1-4, 15-17
A	MIZUUCHI M ET AL.: "Assembly of the active form of the transposase-Mu DNA complex: a critical control point in Mu transposition" CELL, vol. 70, 24 July 1992 (1992-07-24), pages 303-311, XP002129671 figure 7	1-4, 15-17
A	LESCHZINGER A E ET AL.: "Tn552 transposase catalyzes concerted strand transfer in vitro" PROC. NATL. ACAD. SCI. USA, vol. 95, June 1998 (1998-06), pages 7345-7350, XP002129672 figure 1	1-4, 15-17
T	GORYSHIN, IGOR Y. ET AL: "Insertional transposon mutagenesis by electroporation of released Tn5 transposition complexes" NATURE BIOTECHNOLOGY, vol. 18, no. 1, January 2000 (2000-01), pages 97-100, XP000874790 the whole document	1-18

INTERNATIONAL SEARCH REPORT

Information on patent family members

Internat'l Application No

PCT/US 99/21960

Patent document cited in search report	Publication date	Patent family member(s)			Publication date
WO 9810077	A 12-03-1998	US US AU EP PL US	5965443 A 5925545 A 4262097 A 0927258 A 332145 A 5948622 A	A A A A A A	12-10-1999 20-07-1999 26-03-1998 07-07-1999 30-08-1999 07-09-1999

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